

Single-Molecule Biophysics

03/2014 – 06/2014

- Undergraduate research internship

Laboratory of Prof. Sungchul Hohng, Department of Biophysics and Chemical Biology,
Seoul National University

RESEARCH AREAS

- Topics: 1) **Transcriptome-wide post-transcriptional regulation** by RNA-protein interactions and RNA modifications
- 2) **Protein-nucleic acid interactions** for their biological functioning (**CRISPR-Cas9**; telomere and **G-quadruplexes**; synthetic nucleic acids such as BNA and LNA; etc.)
- 3) Cellular dynamics and intracellular signal/force transduction (cellular adhesion, migration, and contraction)
- Spectroscopic techniques: [*in vitro*] **Single-molecule fluorescence spectroscopy** (FRET, PIFE, etc.), Total internal reflection fluorescence (TIRF) microscopy, Alternating laser excitation (ALEX)
[*in cells*] Tension gauge tether (TGT) assays for single-cell mechanobiology
- High-throughput techniques: **Nanopore sequencing**
- Wet lab techniques: [*in vitro*] Site-directed mutagenesis, Gel electrophoresis, Immobilized metal affinity chromatography (IMAC), Fast protein liquid chromatography (FPLC)
[*in cells*] Mammalian cell culture, **Crosslinking immunoprecipitation** (CLIP)
- Dry lab techniques: **MATLAB, Python, IDL, and LabVIEW**

HONORS & AWARDS

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| 2021 | The Best Ph.D. Thesis Award by College of Natural Sciences, Seoul National University |
| 2020 | KAGE Young Researcher Award by Korean Association for Genome Editing (KAGE) |
| 2019 | Best Poster Award by the Committee of East Asian Symposium (EAS) on Single-Molecule Biological Sciences |
| 2018 – 2021 | Global Ph.D. Fellowship by National Research Foundation (NRF) of Korea |
| 2018 | Outstanding Oral Presentation Award by Korean Physical Society (KPS) |
| 2017 – 2018 | Basic Science Fellowship for the Next Generation Research by Seoul National University |
| 2015 – 2016 | Woosan Scholarship for Graduate Students by Woosan Foundation |
| 2011 – 2015 | SNU Tomorrow's Edge Membership (STEM); the Honor Society of Seoul National University by Seoul National University |
| 2009 – 2015 | Presidential Science Scholarship by Korea Student Aid Foundation (KOSAF) |

CONFERENCE PRESENTATIONS

International :

64th Annual Meeting of the Biophysical Society (San Diego, CA, USA; 02/2020),
“Characteristic interactions between BRCA2 and G-quadruplex structures for telomere maintenance”; **Poster**.

2nd East Asian Symposium on Single-Molecule Biological Sciences (Seoul, Korea; 07/2019),
“A regulatory mechanism of CRISPR-Cas9 nuclease specificity revealed from single-molecule structural dynamics”; **Poster [Awarded]**.

63rd Annual Meeting of the Biophysical Society (Baltimore, MD, USA; 03/2019),
“Structural rearrangement of DNA for CRISPR-Cas9 nuclease specificity regulated by the REC2 domain”;
Poster.

2017 SNU-RIKEN Young Investigator Workshop on Molecular Nanospectroscopy (Wako, Japan; 01/2017),
“Structural roles of guide RNAs in the nuclease activity of Cas9 endonuclease”; **Oral Presentation**.

Domestic :

2020 Korean Association for Genome Editing Annual Symposium (Seoul, Korea; 11/2020),
“Mechanisms of Cas9 nuclease specificity revealed by single-molecule analysis”; **Invited Talk as a Laureate of KAGE Young Researcher Award**.

2018 Korean Physical Society Fall Meeting (Changwon, Korea; 10/2018),
“Structural dynamics of DNA for CRISPR-Cas9 nuclease specificity regulated by electrostatic interaction with the REC2 domain”; **Oral Presentation [Awarded]**.

The 122nd General Meeting of the Korean Chemical Society (Daegu, Korea; 10/2018),
“Target specificity of the CRISPR-Cas9 nuclease regulated by the REC2 domain via structural rearrangement of DNA”; **Oral Presentation**.

2017 Korean Physical Society Fall Meeting (Gyeongju, Korea; 10/2017),
“Microscopic mechanism of R-loop expansion for Cas9 nuclease activation”; **Poster**.

The 118th General Meeting of the Korean Chemical Society (Busan, Korea; 10/2016),
“Single-molecule study on guide RNAs as structural regulators for the activation of Cas9 endonuclease”;
Poster.

PATENTS

“Novel Cas9 protein variants with improved target specificity and use thereof”
Patent pending; 31/12/2020; Korea (**K. Sung**, Y. Jung, S. Bae, and S. K. Kim).

PUBLICATIONS

7. J. Lee[†], **K. Sung**[†], S. Y. Joo, S. K. Kim*, and H. Lee* (**†equal contribution**)
“Dynamic interaction of BRCA2 with the telomeric G-quadruplex underlies the telomere replication homeostasis”
To be submitted (2021).
6. S. Y. Bak[†], Y. Jung[†], J. Park[†], **K. Sung**[†], H.-K. Jang, S. Bae*, and S. K. Kim* (**†equal contribution**)
“Quantitative assessment of Cas9 engineering strategies for target specificity enhancement by single-molecule kinetic analysis”
Manuscript under revision (2021).
5. M. H. Jo, B. C. Kim, **K. Sung**, R. Panettieri Jr., S. An*, J. Liu*, and T. Ha*
“Molecular nanomechanical mapping of histamine-induced smooth muscle cell contraction and shortening”
Manuscript submitted (2021).
4. J. Park, **K. Sung**, S. Y. Bak, H. R. Koh*, and S. K. Kim*
“Positive identification of DNA cleavage by CRISPR-Cas9 using pyrene excimer fluorescence to detect a subnanometer structural change”
J. Phys. Chem. Lett. 10, 6208–6212 (2019).
3. **K. Sung**, J. Park, Y. Kim, N. K. Lee, and S. K. Kim*
“Target specificity of Cas9 nuclease via DNA rearrangement regulated by the REC2 domain”
J. Am. Chem. Soc. (Communication) 140, 7778–7781 (2018).
2. C. R. Cromwell, **K. Sung**, J. Park, A. R. Kryslar, J. Jovel, S. K. Kim, and B. P. Hubbard*
“Incorporation of bridged nucleic acids into CRISPR RNAs improves Cas9 endonuclease specificity”
Nature Commun. 9, 1448 (2018).
1. Y. Lim, S. Y. Bak, **K. Sung**, E. Jeong, S. H. Lee, J.-S. Kim, S. Bae*, and S. K. Kim* “Structural roles of guide RNAs in the nuclease activity of Cas9 endonuclease”
Nature Commun. 7, 13350 (2016).